

METHOD OF STERILIZATION

FIELD OF THE INVENTION

This invention relates broadly to sterilization of medical devices. More specifically, this invention relates to a novel apparatus and process for the sterilization of medical devices 5 using ultraviolet radiation.

DESCRIPTION OF THE RELATED ART

Medical device sterilization processes, and in particular commercial contact lens manufacturing sterilization processes, typically involve some form of temperature and/or 10 pressure-based sterilization techniques. For example, a hydrophilic contact lens is typically first formed by injecting a monomer mixture into a mold. The monomer mixture is then polymerized (i.e. the lenses are cured). After other optional processing steps, such as quality inspections, the lens is placed into a container with a solution and the container is sealed. The packaged lens is sterilized by placing the container into an autoclave at an elevated 15 temperature and pressure for an extended period of time, usually at least 15 minutes, typically 30 minutes. Although this commercial process produces thoroughly sterilized contact lenses, the batch-wise autoclave sterilization step is time consuming and costly.

European Patent Application No. 0 222 309 A1 discloses a process using ozone in which packaging material is disinfected in a manufacturing setting. The process involves 20 feeding an oxygen stream into an ozonating chamber, generating ozone from oxygen in the ozonating chamber, placing packaging containers in a sanitizing chamber, feeding the ozone into the sanitizing chamber, and purging the ozone from the sanitizing chamber with sterile air. The process requires that the ozone contact the packaging material for a predetermined time, followed by the sterile air purge step. The process is offered as an alternative to heat- 25 steam sterilization, sterilization by application of electromagnetic radiation, or chemical agent sterilization.

U.S. Patent No. 5,618,492 discloses a process for producing a sterile contact lens in a sealed container during a continuous production process wherein the contact lens is immersed in an ozone-containing solution within a container during a continuous lens packaging 30 process, and the lens and container are subsequently subjected to ultraviolet radiation primarily to degrade the ozone. This process sterilizes the contact lens and the container.

Non-ionizing radiation such as ultraviolet (UV) light is known to damage the DNA of exposed cells. The UV light causes bonds to form thymine dimers which inhibit replication

of DNA during cell reproduction. UV light is used for disinfection in hospital rooms, nurseries, operating rooms and cafeterias. UV light is also used to sterilize vaccines, serum, toxins, municipal waste, and drinking waters. The major weakness of the efficacy of UV light as a sterilizer is that for most materials the radiation is not very penetrating, so the 5 microorganisms to be killed must be directly exposed to the radiation.

A number of patents teach the application of UV light to disinfect and/or inactivate microorganisms to either reduce populations of microorganisms or to eliminate them.

US Patent 5,768,853 and WO96/09775 describe the use of a UV light producing apparatus which deactivates microorganisms in food.

10 US Patent 4,464,336 suggests a method of sterilization by using a flash discharge ultraviolet lamp. The patent teaches that by applying short duration high intensity UV light that microorganisms will be destroyed; however, the conditions for sterilization are not disclosed, nor its application for medical devices.

15 US Patent 5,786,598 discloses the broad concept that a flash lamp system might be used for deactivating microorganisms in containers including a polyolefin container with a foil backing that contains a contact lens and a preservative fluid. Preservation is the use of physical and/or chemical means to kill or prevent the growth of those microorganisms which, by their growth and/or activities, may cause bio-deterioration of a given material or product.

P. Singleton and D. Sainsbury, 1988. Dictionary of Microbiology and Molecular Biology, 20 John Wiley & Sons, New York, NY, pp. 702-703. Although the patent discloses the idea of using a flash lamp system to sterilize contact lenses in a preserved solution in a container, there are no conditions defined to accomplish sterility, nor examples which show that sterility can be accomplished.

25 U.S. Patents 5,034,235 and 4,871,559 disclose the use of intermittent pulses of very intense, very short duration pulses of light to inactivate microorganisms on the surface of food products, and suggests that the method can be used for packages, medical devices, and food products in packages.

30 There still remains a need for a time-efficient, continuous, effective sterilization process for pharmaceutical, medical, and cosmetic products which can be used in an in-line mode in the manufacture of these products.

SUMMARY OF THE INVENTION

This invention provides a process of sterilizing a medical device, and preferably the contents of a sealed container which comprises said medical device, comprising the step of exposing said medical device to ultraviolet radiation whereby the D_{value} of Bacillus

5 stearothermophilus (ATCC 7953) is at least 3.9 milliJoule per square centimeter (mJ/cm^2) ultraviolet radiation in the range of 240-280 nm to the spore. Further, this invention provides a process of sterilizing a medical device comprising the step of subjecting said medical device to ultraviolet radiation wherein the minimum total energy density of said ultraviolet radiation in the range of 240-280 nm which reaches the microorganisms present on said medical device

10 is at least $18 mJ/cm^2$.

This invention further provides an apparatus for delivering pulsed UV radiation to a medical device for sterilization comprising at least one radiation source and a reflector for each said radiation source wherein said at least one reflector directs radiation from said at least one radiation source to a treatment area such that for each pulse of said at least one radiation source, at least $3 J/cm^2$ broad spectrum radiation reaches the treatment area of which at least $45 mJ/cm^2$ of said radiation is UV radiation in the range of 240-280 nm. The treatment area is preferably at the focal plane of said reflector which has the maximum flux, and is where the medical device or container for the medical device is located for treatment.

15 The process and apparatus of the invention is used to provide sterilized medical devices. Further, this invention provides a process and apparatus in which sterilization can be achieved in less than 20 seconds, preferably less than 15 seconds, more preferably in less than 5 seconds. This invention provides a process and apparatus which sterilizes medical devices and optionally sterilizes the contents of the containers holding the medical devices.

20 Preferably the process and apparatus can be incorporated into a manufacturing line. The

25 process and apparatus are efficient and continuous.

DESCRIPTION OF THE FIGURES

The invention will be described with reference to the following figures:

Figures 1 and 2 illustrate one embodiment of the system used in the process of this

30 invention;

Figure 3 shows the sensitivity of Aspergillus niger (ATCC 16404) to radiation of different spectral ranges;

Figure 4 shows the sensitivity of Bacillus stearothermophilus (ATCC 7953) to radiation of different spectral ranges;

Figure 5 shows the sensitivity of Pseudomonas aeruginosa (ATCC 9027) to radiation of different spectral ranges; and

5 Figure 6 shows a graph used to determine the D_{value} of Bacillus stearothermophilus (ATCC 7953).

DESCRIPTION OF THE INVENTION

The term "sterile" or "sterilization" as used herein means the condition of an object, or an environment, which is free of all living cells, all viable spores (and other resistant and disseminative forms), and all viruses and subviral agents capable of replication. Sterility is assured by a minimum sterility assurance level of 10^3 , preferably 10^6 , more preferably 10^9 , and most preferably 10^{12} when the container is inoculated with 10^6 microorganisms. The minimum sterility assurance level is dependent on the type of medical device. For example, for sterilization of a single-use contact lens, the USFDA requires a minimum sterility assurance level of 10^6 in the number of microorganisms per container. A sterility assurance level of 10^6 is the probability of having 1 non-sterile package out of one million packages.

The " D_{value} " is the amount of energy required to kill 90 percent of the organisms present. For steam sterilization, the D_{value} is the time required at a given temperature to reduce the number of viable cells or spores of a given microorganism to 10 percent of the initial number, according to P. Singleton and D. Sainsbury in Dictionary of Microbiology and Molecular Biology, 1988, John Wiley & Sons, New York, NY, pg. 256. According to ANSI standards for gamma radiation, the D_{value} is the radiation dose required to kill 90 percent of the organisms of a homogeneous microbial population, which is defined by assuming that the death of microbes follows first order kinetics. In the case of non-ionizing radiation, the D_{value} will be the non-ionizing radiation dose required to kill 90 percent of the organisms of a homogeneous microbial population, which is defined by assuming that the death of microbes follows first order kinetics. The D_{value} is calculated using the Stumbo-Murphy-Cochran Equation: $D_{value} = U / (\ln N_0 - \ln N_u)$, where N_0 is the initial number of microorganisms in each replicate unit, N_u is $\ln(n/r)$, where n is the total number of replicate units exposed to the sterilizing dose U , and r is the number of units exposed to dose U that test negative for growth. Using the D_{value} , the sterilizing dose can be calculated. For sterilization of a medical

device such as a single-use contact lens, the requirement is for a sterility assurance level of 10^{-6} . The total dose, therefore equals $D_{value} (\log N_o - \log N_u)$.

The term "ultraviolet radiation" or "UV radiation" means radiation having a wavelength or wavelengths between from 200 to 400 nm. If a range is specified following 5 the term "ultraviolet radiation" or "UV radiation", a narrower range of radiation is meant within the 200 to 400 nm range. Further, the range specified, unless otherwise stated, means radiation having a wavelength or wavelengths within the range.

The term "broad spectrum of radiation" means radiation having at least a majority of the wavelengths between 200 to 1100 nm wherein at least a portion of the radiation is UV 10 radiation.

This invention can be used to sterilize medical devices. The configuration of the system used to sterilize the medical device depends on the transmissiveness of the medical device to UV radiation (240-280 nm). If the medical device is transmissive to at least a portion of the ultraviolet radiation (240-280 nm), for example, preferably greater than 10 percent, more preferably more than 50 percent, most preferably more than 75 percent, then a single radiation source can be used to sterilize the medical device as long as at least 18 15 mJ/cm², more preferably at least 30 mJ/cm², most preferably at least 36 mJ/cm² of UV radiation in the range of 240-280 nm reaches all the microorganisms and/or all surfaces of the medical device to be sterilized. If the medical device is not transmissive to UV radiation 20 (240-280 nm) or is transmissive to such a small percentage of UV radiation (240-280 nm), for example, less than 10 percent, then more than one radiation source will most likely be necessary to sterilize the medical device. However, any configuration and any number of radiation sources can be used as long as the minimum levels of energy specified herein reach all the microorganisms or all the surfaces of the medical device which are to be sterilized.

25 This invention is preferably used to sterilize medical devices which are in sealed containers. If the medical device is to be sterilized after placing it in its container, the container must be at least partially transmissive to ultraviolet radiation (240-280 nm), preferably the container is transmissive to at least 25% of the ultraviolet radiation (240-280 nm), more preferably the container is transmissive to at least 50% of the ultraviolet radiation 30 (240-280 nm), and most preferably the container is transmissive to at least 60% of the ultraviolet radiation (240-280 nm). Ideally, the container is transmissive to substantially all of the ultraviolet radiation (240-180 nm). If the medical device is transmissive to at least a portion of the ultraviolet radiation (240-280 nm), and a single radiation source is used to

sterilize the medical device, and if the medical device is in a container, then the container can be transmissive to at least a portion of UV radiation only in one area of the container, as long as enough radiation can reach all the microorganisms and all of the surfaces of the medical device and contents of the container. (The contents of the container include the inside 5 surfaces of the container and any solution or other storage medium for the medical device which is inside the container). However, if more than one radiation source is used, it is preferred that the container is at least partially transmissive to ultraviolet radiation (240-280 nm) over most of the surface area of the container, and preferably substantially over the entire surface area, that is, that the container, at the time of exposure to the sterilizing radiation, 10 preferably does not comprise any materials that have less than 10 percent, more preferably not less than 25 percent ultraviolet radiation (240-280 nm) transmissivity. More preferably, the container is transmissive in substantially all directions to at least 50% of the UV radiation (240-280 nm), and most preferably the container is transmissive in substantially all directions to at least 60% of the UV radiation (240-280 nm). Ideally, the container is transmissive in 15 substantially all directions to substantially all of the UV radiation (240-280 nm).

Examples of medical devices which may be used in the process of this invention include, for example, catheters, surgical equipment, implants, stents, sutures, packing, staples, and bandages, and the like. Materials which may be used to make the medical devices include metals, glycerol monomethacrylate, polyvinylalcohol, polyvinylpyrrolidone, 2- 20 hydroxyethyl methacrylate (HEMA), methacryloxypropyltris(trimethylsiloxy)silane, polydimethylsiloxane, methylacrylic acid, methylmethacrylate, urethanes, polypropylene, polylactide, polyglactide, polyethylene glycol, polypropylene glycol, and the like, and the materials described below.

To decrease the complexity and energy demands of this method of sterilization, it 25 would follow that it would be preferred that the medical device is at least partially transmissive to ultraviolet radiation, preferably the medical device is transmissive to at least 10% of the ultraviolet radiation, more preferably the medical device is transmissive to at least 25% of the ultraviolet radiation, and most preferably the medical device is transmissive to at least 50% of the ultraviolet radiation. The preferred medical device treated by the method of 30 this invention is a contact lens. It is more preferred that the contact lens is in a hermetically sealed contact lens container, and even more preferred that the contact lens container holds a liquid in which the contact lens is immersed. Even though the method would be simplified by using a UV transmissive medical device, the presently preferred contact lens is a contact

lens comprising a UV-blocker which blocks greater than 30%, more preferably greater than 50%, and most preferably greater than 80% of the UV radiation (240-280 nm) impinging upon it. The preferred embodiment, the process of sterilizing a contact lens, will be described herein; however, it is apparent that other medical devices, such as those listed above, can be substituted for the contact lenses in the method described in detail below for contact lenses.

It is preferred that the process of this invention is incorporated into an in-line continuous contact lens manufacturing and packaging process. In the most preferred embodiment, the contact lens is formed and placed in a portion of a contact lens container, solution is added to the container, the container is sealed and the container is subjected to short duration, high intensity pulses of radiation, including ultraviolet radiation (240-280 nm) to produce a sterile packaged contact lens which is ready for distribution and use. In the preferred embodiment, the contact lens comprises a hydrogel material and the contact lens is stored in an aqueous solution in the container. The manufacture and placement of the contact lens in the container can be by any process for making contact lenses including those described in, for example, US Patents 5,435,943; 5,395,558; 5,039,459; 4,889,664; 4,565,348; and 4,495,313, incorporated herein by reference. Other methods are disclosed in other patents, and are known to a person of ordinary skill in the art.

The preferred method provides that a contact lens mold is formed by injection molding two thermoplastic contact lens mold halves which, when put together, form a cavity in the shape of the contact lens. These thermoplastic contact lens molds are typically used once to form a contact lens. Reusable lens molds made out of more durable materials, for example, glass or metal can also be used. Typically, before the lens molds are put together, the monomer or prepolymer mixture which forms the contact lens polymer is injected into a first mold half and a second mold half is placed onto the first mold half which pushes out any excess monomer or prepolymer mixture. However, a one-piece mold can be used to form the contact lens, or the monomer or prepolymer mixture can be injected between the molds after assembly of the molds. The monomer or prepolymer mixture is then cured to form the contact lens. Curing of the monomer or prepolymer mixture is preferably initiated by using photoinitiation. After curing the monomer or prepolymer mixture, the mold halves are removed, and the contact lens is hydrated, if needed. After hydration, preferably one contact lens is placed in a contact lens container. It is preferred that each contact lens container also contains at least enough aqueous solution to fully wet the contact lens in the container, but the presence of aqueous solution is not required in the process of this invention. The aqueous

solution, if present, can be added to the container before or after placing the contact lens in the container. Further, the container can be sealed before or after the sterilization step.

Alternatively, the container can be sealed before and after the sterilization step if multiple layers of the container are added in multiple steps. For example, a sealed container can be

5 shrink-wrapped or a label added after the sterilization step; or there can be multiple sterilization steps before and after sealing the container. For example, the container can undergo the sterilization step while empty, and/or when it has liquid in it and/or when it has a contact lens placed in it and is not sealed, and/or after the container is sealed with the contact lens and optional liquid present in the container. In additional embodiments, the individual
10 parts and contents of the container can be individually treated by the method of this invention, and then assembled, and/or treated again. It is presently preferred to hermetically seal the container in which a contact lens is present prior to the process of sterilization, and to perform the step of subjecting the hermetically sealed container having the contact lens therein to UV radiation (240-280 nm) only once. The container can be sealed by any means which provides
15 a hermetically sealed container.

The contact lenses useful in accordance with the present invention may be formed from any materials useful for contact lenses. For example, the lenses may be hydrophilic lenses formed from the polymerization or copolymerization of acrylates or methacrylates, such as 2-hydroxyethyl methacrylate (i.e., HEMA); hydrophobic lenses formed from
20 polysiloxanes; or lenses formed from copolymers displaying a range of hydrophobic and hydrophilic properties. The preferred contact lens material is Etafilcon-A which comprises HEMA, MAA, EGDMA, TGDMA, and Darocur. Useful contact lens materials are described in US Patents 5,484,863; 5,039,459; 4,889,664; 5,0684,058; 5,654,350; 5,648,402; 5,311,223; 5,304,584; 5,256,751; 5,196,458; 4,495,313; and 4,680,336, incorporated herein
25 by reference.

Additionally, the process of this invention can be used to sterilize contact lenses which contain ultraviolet radiation blocking agents. Contact lenses containing ultraviolet radiation blocking agents include, Acuvue® and Surevue® made by Johnson & Johnson Vision Products. Blocking agents which can be used in contact lens compositions include
30 Norbloc™ 7966, which is 2-(2'-hydroxy-5-methacryloxyethylphenyl)-2H-benzotriazole; other benzotriazoles, such as, 2-(2'-hydroxy-5'-methacryloxyethylphenyl)-2H-benzotriazole; 2-{3'-tert-butyl-2'-hydroxy-5'-(3"methacryloyloxypropyl)phenyl}-5-chlorobenzotriazole; and benzophenones, such as, 1,3-bis(4-benzoyl-3-hydroxyphenoxy)-2-

propyl)(meth)acrylate; 2-hydroxy-4-((meth)acryloxyethoxy)benzophenone; 4-methacryloxy-2-hydroxybenzophenone, others disclosed in US Patents 5,133,745; 4,528,311; 4,716,234; 4,528,311 and 5,681,871, and others known to a person of ordinary skill in the art. Contact lenses can be made according to the examples and teachings in US Patents 5,133,745; 5 4,716,234; 4,528,311; 4,304,895 and 5,681,871. Also the process of this invention can be used to sterilize contact lenses which contain tints, including visibility tints, including Reactive Blue #4 (2-anthracenesulfonic acid, 1-amino-4-(3-((4,6-dichloro-s-triazin-2-YL)amino)-4-sulfoanilino)9,10-dihydro-9,10-dioxo-) and other materials disclosed in U.S. Patent 5,292,350. This method can also be used to sterilize opaque lenses and lenses 10 comprising titanium dioxide. The most preferred lens material is Etafilcon-A further comprising NorblocTM and Reactive Blue #4.

The aqueous solution, if present in the contact lens container, is preferably buffered to a pH of between about 6.5 to about 7.8 in order to approximate the pH of the fluids in the eye. The solution may be buffered by a wide variety of buffers including phosphates, borates, 15 citrates, and other biocompatible buffers known to a person of ordinary skill in the art. The presently preferred buffer is a borate solution. The amount of solution depends on the size of the container. The solution preferably does not contain any preservatives. Typically, the container has a volume between 0.5 milliliters and 50 milliliters, preferably about 1 milliliter, and there is between 0.1 milliliter to 1 milliliters, preferably about 0.5 milliliters of solution 20 in the container.

Containers which are useful in this invention are any of the known containers which are or can be hermetically sealed as long as the containers are at least partially transmissive to ultraviolet radiation (240-280 nm). The containers can be UV transmissive glass, thermoplastic pouches and bags, cyclic olefin copolymers, injection molded or thermoformed 25 plastic containers, and conventional bowls and lids for contact lenses, as long as enough ultraviolet radiation (240-280 nm) can penetrate the container to sterilize the contents of the container. It is presently preferred that the contact lens container comprises a bowl and a lid. It is preferred that the material or materials of at least the bowl of the container are at least 30 partially transmissive to ultraviolet radiation. Particularly, for the sterilization of a contact lens comprising UV-blocker, it is even more preferred that the bowl and the lid are at least partially transmissive to ultraviolet radiation, preferably in all directions. To accomplish this, it is preferred to replace the conventional foil lid with a thermoplastic lid, which may consist of one or more layers of, for example, polyolefins, such as, polyethylenes, polypropylenes,

polybutylenes, and copolymers of the above; cycloolefins (COC); halogenated films, such as polyvinylchlorides (PVC), polyvinylidine chlorides (PVDC), polyvinylidene fluorides, polymonochlorotrifluoroethylenes (PCTFE), polyvinylidene fluorides (PVDF), and polyfluorocarbons; polyurethanes; polyamides; polyimides; ethylene-vinyl acetate

5 copolymers (EVA); ethylene vinyl alcohols (EVOH); ethylene acrylic acid copolymers (EAA); acrylics, such as polymethylmethacrylates; ionomers; and cellulose materials, such as cellulose esters, and cellophanes. It is presently preferred that the bowl is a polyolefin, and the lid is a multilayered structure comprising polypropylene. The materials of the bowl and the lidstock should preferably be free of any component that will scatter light. The most
10 preferred method of sealing the container is to heat seal the thermoplastic lid to the thermoplastic bowl. The most preferred containers and materials for the container are described in a simultaneously filed application by James Peck, et al, U.S. Serial
A No. 69/259,795 entitled "Package for Medical Device" (VTN-445) which is incorporated
herein by reference.

15 Sterilization of the contact lens and preferably the contents of the container is achieved by subjecting the container to short duration, high intensity radiation comprising ultraviolet radiation (240-280 nm), wherein the energy density of said high intensity ultraviolet radiation (240-280 nm) inside said container is sufficient to provide complete inactivation of the respective microorganism. The radiation must comprise UV radiation (240-280 nm), because it has been determined that radiation in the ultraviolet radiation range is most responsible for the inactivation of microorganisms; however, the IR, and VIS areas of the spectrum may or may not contribute to the inactivation depending on the microorganism. Different microorganisms are known to be affected differently by ultraviolet radiation (240-280 nm). Viruses are susceptible to UV radiation, and vegetative bacteria are more
20 susceptible to UV radiation. The spore-forming microorganisms are known to be the most resistant to UV radiation. The reasons for the resistance of the spores to UV radiation is primarily attributed to the composition of the outer coat of the spore and the ability of spores to refract light. Therefore, a range of intensities will affect different types of microorganisms differently. We have determined that one of the most resistant microorganism to this method
25 of sterilization is Bacillus stearothermophilus (ATCC 7953). The D_{value} for Bacillus stearothermophilus (ATCC 7953) is at least 3.9 mJ/cm² ultraviolet radiation (240-280 nm) to the spores. For a sterility assurance level of 10⁻³, the dose to the microorganisms (where the initial inoculum is 10⁴ cfu/container) is at least 30 mJ/cm² of UV radiation (240-280nm). For
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a sterility assurance level of 10^{-6} , the dose to the microorganisms is at least 41 mJ/cm^2 of UV radiation (240-280nm). For a sterility assurance level of 10^{-9} , the dose to the microorganisms is at least 52 mJ/cm^2 of UV radiation (240-280nm). For a sterility assurance level of 10^{-12} , the dose to the microorganisms is at least 65 mJ/cm^2 of UV radiation (240-280nm). This energy
5 is preferably supplied in a single flash by a flash lamp. For an initial inoculum of 10^6 , the microorganisms are exposed to at least 45 mJ/cm^2 of said UV radiation (240-280nm) to provide a sterility assurance level of 10^{-6} . In the preferred embodiment, in which the medical device is a UV-blocker lens in a container having approximately 50% transmissivity for UV radiation (240-280 nm), the energy is provided by two flash lamps each having a reflector and
10 each of which provides approximately 3.5 J/cm^2 broad spectrum of which 50 mJ/cm^2 is UV radiation (240-280 nm) to the container, and the lamps flash simultaneously, whereby the contact lens, and more preferably the contact lens and the contents of the container are rendered sterile. The container permits approximately 50 percent of the UV radiation impinging on it from each lamp to reach the contents of the container.

15 The high intensity ultraviolet radiation can be generated and directed to the container by any means including lasers, and lamps, with or without reflectors, lightguides, lightpipes, fiber optics, dioptric or catadioptric optical systems to focus light on the product. It is preferred that at least 10%, more preferred at least 30% and most preferred at least 50% or more of the total energy which reaches all the surfaces of the medical device to be sterilized is ultraviolet radiation (240-280 nm). For a medical device in a container, it is preferred that at
20 least 10%, more preferred at least 30% and most preferred at least 50% or more of the total energy which reaches all the surfaces of the medical device, and the contents of the container is UV radiation (240-280 nm). The high intensity ultraviolet radiation, generated by any means including lasers, and lamps, pulsed or continuous, can be adjusted by the use of filters,
25 choppers, splitters, optics, dichroic mirrors, diffractive mirrors, gratings, bandpass filters, short pass filters, long pass filters, dielectric coatings, multilayer coatings, selective beamsplitters, monochromators, Fabry-Perot Etalon, reflective coatings, binary optics, acoustic modulators, or prismatic devices to focus the radiation on the container, or to limit the radiation which reaches the medical device or the container to the germicidally effective
30 wavelengths, or to eliminate radiation at wavelengths which will detrimentally impact the container polymer or medical device polymer. For many polymers which are commonly used for contact lens containers and contact lenses, which were described earlier, radiation at wavelengths less than 320 nm is absorbed by the polymers and may cause chain scissions

within the polymers. To protect the polymers from degradation, the means to adjust the wavelengths of the radiation, which reaches the contact lens container, can be present between the radiation source and the contact lens container, or the container can be constructed to filter out certain regions of the spectrum.

5 Surprisingly, it was found that the presence of UV-blocker agents in the contact lens polymer provides protection to the contact lens polymer. For example, an Etafilcon-A contact lens without UV blocker can withstand up to approximately 35 mJ/cm² UV radiation (240-280 nm) and/or approximately 5 J/cm² broad spectrum radiation delivered to each side of the lens by each of two flash lamps which flash simultaneously in multiple flashes before

10 its mean equilibrium water content of the contact lens is increased such that it falls out of specification; however, an Etafilcon-A contact lens comprising a UV blocker, such as, NorblocTM can withstand more than 150 mJ/cm² UV radiation (240-280 nm), and/or approximately 10 J/cm² broad spectrum radiation delivered to the contact lens before its mean equilibrium water content is increased such that it falls out of specification.

15 The sterilization dose of radiation can consist of only ultraviolet radiation having wavelengths of 200 to 400 nm, or between from 240 to 280 nm, or radiation at a single wavelength, or the radiation can consist of a broad spectrum of wavelengths including visible, infrared, or any combination of the above, as long as at least a portion of the spectrum is at 254 nm or between from 250 to 260 nm. In the presently preferred embodiment, each flash lamp produces 3.5 J/cm² broad radiation of which approximately 11.9% of the total energy is between from 200 to 400 nm, 1.8% is between from 240 to 280 nm, and 0.5% is between from 250 to 260 nm. A more preferred radiation source would be one in which at least 25% of the total energy is between from 240 to 280 nm; more preferably at least 50%, and most preferably greater than 90% of the total energy is between from 240 to 280 nm.

20 25 Sterilization of the container, which preferably comprises at least one contact lens and aqueous solution is preferably achieved by subjecting the contact lens and container to short duration, high intensity ultraviolet radiation, wherein the energy density of said high intensity ultraviolet radiation in the range of 240-280 nm at all the surfaces of the contact lens is at least 18 mJ/cm², more preferably at least 30 mJ/cm², most preferably at least 36 mJ/cm², whereby the contact lens, and more preferably the contact lens and the contents of the

30 container are rendered sterile.

Note that the monitoring system that was used to measure all the spectroradiometric energies reported herein is further described in Ebel, et al, US Patent Application

A 69/259,796, (VTN-443) entitled "Sterilization System", filed concurrently herewith, and incorporated herein by reference. The monitoring system disclosed in Ebel et al, can measure differences in the spectroradiometric output of each flash, even if the total energy level of the flash does not change. That application indicates in an example the importance of measuring 5 the spectroradiometric radiation, because in the preferred embodiment, a measurement of the broad spectrum radiation does not indicate if the amount of UV radiation (240-280 nm) as a portion of the total radiation has dropped below the necessary amount to achieve sterilization.

In the preferred embodiment, the ultraviolet radiation is delivered to the container by a system which pulses or flashes the container with a broad spectrum of radiation including 10 ultraviolet radiation. The pulses or flashes are short and intense, that is, the pulses last less than 5 seconds, more preferably less than 3 seconds, and most preferably less than 1 second. In the preferred system each flash lasts less than 0.5 second, more preferably less than 0.1 second, most preferably about 1 millisecond, and the time between flashes is about 100 to about 200 milliseconds; therefore, the sterilization process can be complete within seconds 15 even if it takes multiple flashes or pulses to deliver the sterilizing dose. The pulses or flashes deliver enough energy to sterilize the medical device and contents of the container. Unlike continuous ultraviolet radiation which delivers a uniform amount of energy over a period of time, each flash or pulse of energy in this system delivers a large amount of energy to surfaces of the contact lens and the container in a short period of time. It is preferred that at 20 least 18 mJ/cm² (240-280nm) reaches the surface of the contact lens in less than 10 milliseconds, more preferably in less than 1 millisecond, most preferably in less than 500 microseconds. More preferably, at least 30 mJ/cm² reaches the microorganisms in less than 1 millisecond. It has been found that the effectiveness of the system at sterilizing is dependent 25 not only on the total amount of energy that is supplied to the container, but that there is a maximum amount of time (or flashes) in which that energy needs to be delivered to the container. It has been determined that the energy required to render the contact lens sterile is preferably delivered to the surface of the contact lens in at most three flashes, more preferably at most two flashes and most preferably in one flash of at least one radiation source in a pulsed UV radiation system. It is preferred that the flash lamp apparatus or each flash lamp 30 in the preferred embodiment provides at least 20 mJ/cm², more preferably at least 25 mJ/cm², and most preferably at least 30 mJ/cm² UV radiation (240-280 nm) in each flash to the microorganisms. It is also preferred that the flash lamp apparatus or each flash lamp in the preferred embodiment produces at least 40 mJ/cm², more preferably at least 50 mJ/cm² and

most preferably at least 60 mJ/cm² UV radiation (240-280 nm) in each flash. For a medical device in a container, the medical device having less than 20 percent transmissivity (240-280 nm), it is preferred that multiple radiation sources flash simultaneously to produce at least 80 mJ/cm² total UV radiation (240-280 nm) per flash, more preferably at least 100 mJ/cm² total UV radiation (240-280 nm) per flash, and most preferably at least 120 mJ/cm² total UV radiation (240-280 nm) per flash which impinges on the container.

Additional benefits of a pulsed ultraviolet radiation system over a continuous ultraviolet radiation system is that it generates less heat, and that the system can be monitored to verify that a suitable spectrum was generated for every flash, using the monitoring system disclosed in Ebel et al., U.S. Patent Application 691,259,796 (VTN-443) entitled "Sterilization System", earlier incorporated herein.

The application of the short, high intensity ultraviolet radiation can be accomplished by using one or more sources of radiation. If one source of radiation is used it is preferably directed at the medical device at the base, preferably the bowl, of the container. If two sources of radiation are used, preferably one is directed at the medical device through the base and the other is directed through at the top, preferably at the lid of the container. As long as the contents of the container receives a sterilizing dose of radiation, and each single cell of microorganism is exposed to sufficient energy to inactivate it, the configuration of the sources of the radiation is not important. Only one radiation source is necessary when the container and medical device have sufficient transmissivity to allow sufficient energy to transmit through the container and device to affect sterility on the entire contents of the container. For example, one radiation source may be effective for a container holding a contact lens which does not have an UV-blocker in its polymer, or a contact lens that is transmissive to greater than 30 %, more preferably greater than 50 % of UV radiation (240-280 nm).

The presently preferred apparatus for delivering the high intensity radiation to the contact lens having a UV-blocker in a container with an aqueous solution in the container is a PurePulse PBS1-4 system (manufactured by PurePulse Technologies, Inc., San Diego, CA) which consists of two flash lamp assemblies connected in series, each of which consists of a lamp, and a reflector in a housing. The PBS1-4 system further consists of a pulse-generator capable of generating a large pulse of energy by virtue of its large capacitance (80-160 µF) and high potential (greater than 6 kV), and control circuits. Each lamp generates a broad spectrum of radiation which includes ultraviolet, infrared and visible light. The system is shown in Figures 1 and 2. Figure 1 shows two lamp assemblies 35 and 36. Lamp assembly

35 comprises lamp 21, reflector 23 and lamp housing 25, having a protective quartz plate 39. Lamp assembly 36 comprises lamp 22, reflector 24 and lamp housing 26, having a protective quartz plate 38. Also shown in Figure 1 is the chamber 40, which was added to the PBS1-4 system. The chamber comprises a drawer front 27, the product support 28, and spacer 29.

5 The spacer 29 separates and supports the lamp assemblies and moveably holds the product support 28 via brackets 41 and 43 at a fixed distance from the lamp assemblies. The product support 28 is attached to the drawer front 27. Each flash lamp 21 and 22 contains a rare gas as a luminous component, preferably xenon, within a glass envelope or cylinder, preferably made of quartz, more preferably made of sapphire. The lamps comprising sapphire are preferred, because they can withstand higher internal pressures over a range of temperatures, produce less devitrification and have higher transmission coefficients over a broad range of wavelengths. The flash lamps are commercially available, for example from ILC Technology, Inc., Sunnyvale, CA. The lamp assemblies 35 and 36 face each other with a space between the lamps for the product support 28 holding the container or containers to be sterilized.

10 The system is shown with one container 30 on the product support 28 ready for sterilization. The flash lamps flash substantially simultaneously, preferably within less than 5 microseconds, each generating a minimum of 40 mJ/cm^2 of UV radiation (240-280 nm), more preferably a minimum of 50 mJ/cm^2 (240-280 nm) per flash. The system is light-tight when the lamps flash, due to the chamber 40 which is rendered light-tight when the product support

15 28 is pushed into the space between the lamp assemblies 35 and 36 and the drawer front 27 creates a light-tight seal with the front of spacer 29. The system has at least one capacitor (not shown) which stores the energy at a high voltage needed to cause both lamps to flash substantially simultaneously. Each flash lamp 21 and 22 is at least partly surrounded by a reflector 23 and 24 which focuses light in the direction of the container 30. The container 30,

20 25 comprises a contact lens 37 and aqueous solution (not shown). Preferably, the reflectors 23 and 24 have a cylindrical, polygonal, parabolic or elliptical shape and a semi-circular cross-section around the flash lamps. The reflectors are formed by extruding aluminum and adding reflective coatings. More information on flash lamp systems can be found in US Patents 4,464,336; 5,034,235; and 4,871,559, incorporated herein by reference.

30 The containers are preferably placed between the two lamps so that the contact lenses are closest to the center-line between the two lamps, and in the focal planes of the reflectors. Typically, multiple contact lens containers will be inserted between the lamps and flashed together to render the multiple containers of the contact lenses sterile simultaneously. If two

rows of containers are to be flashed simultaneously, they are positioned in the center between the lamps facing bowl 31 to bowl with the tabs 32 facing outward. Preferably, 6 or more containers are flashed simultaneously, more preferably, 8 or more containers, most preferably 10 or more containers are flashed simultaneously. The presently preferred embodiment flashes 12 containers some of which can be removably-attached to each other. These removably-attached containers are commonly used to configure multi-packs of contact lens containers.

The contact lens containers can be inserted and removed from between the flash lamps by any means. Presently it is preferred that the lamps face each other with one facing in the direction of the floor and the other facing in the direction of the ceiling. Further, it is presently preferred that the lamps remain stationary and that a moveable product support 28 which behaves like a drawer is used to move the contact lens containers into and then out of the space between the lamps. The product support 28 which holds the one or more contact lens containers between the two lamps preferably comprises a quartz glass tray. Fused silica, synthetic fused silica, or sapphire may be substituted for the quartz glass. In operation, the tray is first loaded with the one or more containers to be flashed, the tray and attached drawer front are pushed into the support between the lamps which renders the chamber light-tight, the lamps are flashed, and after the containers have received the appropriate dose, delivered in at least one flash, the sterilized contact lens containers are removed from the tray. The containers are preferably placed on the product support with the bowl of the container closest to the flash lamp which faces the ceiling and the lid of the container closest to the flash lamp which faces the floor.

Alternatively, the product support can consist of any materials which will hold the containers and permit enough ultraviolet radiation (240-280 nm) to reach the contents of the one or more containers. For example, it can comprise polymeric or other glass materials in any configuration which holds the containers, e.g. sheet, mesh, or bars. It is presently preferred that the product support comprises quartz. Preferably, only when a light-tight chamber is established will the lamps be permitted to flash. A light-tight chamber will prevent any photo-annealing of the microorganisms' damaged DNA after subjecting the medical device to UV radiation (240-280 nm).

Alternatively, the lamps can be positioned facing in any direction. Preferably the lamps face each other. For example the lamps can be positioned facing opposite walls. The containers can either be stacked up within the chamber or more preferably an array of

removably-attached containers can be hung between the lamps, for example by a hook, clasp, or clip, or the containers can be moved into the space between the lamps by a conveyor.

In the preferred embodiment, the distance between the lamp assemblies is determined by the focal planes of the reflectors. In the present embodiment, the surfaces of the container are 21 mm away from the lamp assemblies. Preferably the reflector is designed to minimize the working distance between the lamp assemblies, provided there is enough space between the lamp assemblies for insertion and removal of the containers. For the preferred embodiment, the reflector is preferably designed to focus at least 50 mJ/cm² UV radiation (240-280nm) and/or at least 3.5 J/cm² broad spectrum at the containers.

The energy levels specified herein can be used to determine the number of flashes which are needed to sterilize a medical device in a container. The transmissivity of a container and the medical device within the container must be determined. As taught herein, it is now known that the minimum level of energy which must reach the inside of the container and the surface of the medical device to obtain sterilization must be equivalent to at least 18 mJ/cm² of UV radiation in the range of 240-280 nm. Then, the following formula with reference to Figure 1 can be used to calculate the amount of energy. The following formula is suited for a system which has two energy sources, but can be modified if one or more than two energy sources are used:

Total energy from all sources $E(\lambda) = E_a(\lambda) + E_b(\lambda)$, where $E_a(\lambda)$ indicated in Figure 1 is the energy above the contact lens 37 in the container 30, and $E_b(\lambda)$ indicated in Figure 1 is the energy below the contact lens 37 in the container 30.

$$\text{where } E_a(\lambda) = E_U(\lambda) [e^{-k(\lambda)_{LS} x_{LS}}] + E_L(\lambda) [e^{-k(\lambda)_{B B} x_B}] [e^{-k(\lambda)_{L L} x_L}],$$

$$\text{and } E_b(\lambda) = E_U(\lambda) [e^{-k(\lambda)_{LS} x_{LS}}] [e^{-k(\lambda)_{L L} x_L}] + E_L(\lambda) [e^{-k(\lambda)_{B B} x_B}]$$

where transmissivity $T = I$ (intensity which penetrates a material)/ I_0 (intensity incident on a material) = $e^{-k(\lambda)x}$ where $k(\lambda)$ is the transmissivity constant of a material and x is the thickness of the material. $E_U(\lambda)$ is the energy from the upper lamp indicated in Figure 1 by the arrow labeled $E_U(\lambda)$. $E_L(\lambda)$ is the energy from the lower lamp indicated in Figure 1 by the arrow labeled $E_L(\lambda)$. The subscript LS indicates lidstock. The subscript L indicates lens. The subscript B indicates bowl. $E_a(\lambda)$ and $E_b(\lambda)$ each have to be at least 18 mJ/cm² UV radiation (240-280 nm) to achieve sterilization of the medical device. If the transmissivity of the container or medical device is increased or decreased, the energy of the radiation sources can be calculated, because $E_a(\lambda)$ and $E_b(\lambda)$ are known. (For the preferred embodiment herein, the contribution of the UV radiation (240-280 nm) which passes through the UV-blocking

contact lens was assumed to be zero when determining the amount of UV radiation which the microorganism is subjected to on each side of the lens.)

Initially, a prototype system consisting of two lamps was evaluated capable of delivering 1.5 J/cm² broad spectrum from each lamp to the outside of the container. Since the 5 ultraviolet radiation (240-280 nm) is the recognized vehicle of kill of the microorganism, it was necessary to quantify the output of these lamps for ultraviolet radiation (240-280 nm). Extensive lamp mapping studies showed the output of each lamp at 21 mm from the bottom of the lamp housing to be on average 25 mJ/cm² (240-280 nm) from each of the lamps to the containers for an area 60 mm wide by 180 mm long. A succession of microbiological tests on 10 a battery of microorganisms were conducted to determine if sterility could be achieved.

Microbiological evaluation of the effectiveness of the system was conducted using containers, consisting of bowls and lidstock, (both about 50 percent transmissible to UV radiation (240-280 nm)), holding UV-blocking contact lenses (20% transmissive to 240-280 nm) in a non-preserved solution of buffered borate. The contact lens containers were made 15 according to Example 1 in "Package for Medical Device" (VTN-445), U.S. Serial No.

A 69/259,795, earlier incorporated herein. The test microorganism was added at a concentration of 10⁴ colony forming unit/package (cfu/pkg). The closed intact containers were centered between the lamps and simultaneously subjected to 25 mJ/cm² (240-280 nm) from each lamp. One hundred containers containing Aspergillus niger (ATCC 16404), 20 Bacillus stearothermophilus (ATCC 7953) spores or Bacillus subtilis (ATCC 9372) spores were exposed to 1-flash for a total of 50 mJ/cm² UV radiation (240-280 nm) (25 mJ/cm² per lamp) from both the top and bottom lamps. The containers inoculated with Aspergillus niger (ATCC 16404) were then processed in a laminar flow hood whereby the entire contents of the container were placed into potato dextrose broth and incubated at 25° C for 14 days. The 25 containers inoculated with Bacillus spore preparations were transferred to tubes containing 40 ml of trypticase soy broth and incubated for 14-days at 35-37°C. This tube terminal sterilization method allows for the detection of the viability of 1-single cell. After 14-days of incubation, the tubes were visually evaluated for turbidity and designated as positive for growth or negative for no growth. The positive tubes were subsequently identified and 30 confirmed as the microorganism inoculated in the test. This experiment was repeated with 100 additional tubes for 2 flashes (50 mJ/cm² cumulative exposure from each lamp (240-280 nm)) both lamps flashing simultaneously, and repeated again for 3 flashes (75 mJ/cm² cumulative exposure from each lamp (240-280 nm)) both lamps flashing simultaneously.

The number of test tubes with viable test microorganisms out of the one hundred tested at each energy level were recorded in Table 1.

The results in Table 1 clearly show that the amount of energy produced by the system was capable of inactivating some of the test microorganism; however, it was not effective in inactivating any of the more resistant spore formers. However, this system could be used to deliver enough energy in one flash to sterilize a non-UV-blocking contact lens in an aqueous solution in a container having transmissivity greater than 50% for 240-280 nm radiation.

TABLE 1 Number of positive tubes/100

Microorganism	1 flash *	2 flashes *	3 flashes *
<u>Aspergillus niger</u> (ATCC 16404)	2	1	4
<u>Bacillus stearothermophilus</u> (ATCC 7953)	89	68	4
<u>Bacillus subtilis</u> (ATCC 9372)	100	81	29

* each flash delivering 25 mJ/cm² (240-280 nm) per lamp

A series of modifications, including an increase in the capacitance of the power supply in the pulse generator, increased the full spectrum output of each lamp to a minimum of 3.5 J/cm² with the UV radiation output of 50 mJ/cm² (240-280 nm) over the same 60 mm x 180 mm treatment area. In this system the two lamps flashed simultaneously. A microbiological test of the output showed that increasing the UV-radiation output from 25 mJ/cm² to 50 mJ/cm² in the spectral region from 240 – 280 nm of each lamp resulted in sufficient UV radiation to sterilize Aspergillus niger (ATCC 16404), Bacillus cereus (an environmental isolate), Bacillus subtilis (ATCC 9372), Candida albicans (ATCC 10231), Pseudomonas aeruginosa (ATCC 9027), Staphylococcus aureus (ATCC 6538) and Serratia marcescens (ATCC 13880). The results of this test are listed in Table 2. Each cell in Table 2 indicates the number of containers out of 100 containers which tested positive, that is, which had viable test microorganisms in them.

TABLE 2

Microorganism	1 flash *	2 flashes *	3 flashes *
<u>Aspergillus niger</u> (ATCC 16404)	0	0	0
	0	0	0
	0	0	0
<u>Bacillus cereus</u>	0	0	0
<u>Bacillus stearo-</u> <u>thermophilus</u> (ATCC 7953)	0	0	0
	0	0	0
	0	0	0
<u>Bacillus subtilis</u> (ATCC 9372)	1	0	0
	1	0	0
	1	0	0
<u>Candida albicans</u> (ATCC 10231)	0	0	0
	0	0	0
	0	0	0
<u>Pseudomonas</u> <u>aeruginosa</u> (ATCC 9027)	0	0	0
	0	0	0
	0	0	0
<u>Staphylococcus</u> <u>aureus</u> (ATCC 6538)	0	0	0
	0	0	0
	0	0	0
<u>Serratia marcescens</u> (ATCC 13880)	0	0	0
	0	0	0
	0	0	0

* each flash delivering 50 mJ/cm² (240-280 nm) per lamp

While the modified machine proved capable of delivering the required amount of
 5 energy, the modifications reduced the lamp life from an expected 20-30 million flashes to 2-5
 thousand. In manufacturing, it is not practical to consider running the machine with this
 reduced lamp life; therefore, the machine requires further modifications to extend the lamp
 life. We have determined that the reflectors need to be redesigned to focus the light energy
 from the existing lamps to the container to a smaller area of 25 mm x 180 mm, thus
 10 increasing the fluence of each reflector at the focal plane of the reflector to a minimum of 3.5
 J/cm² of broad spectrum radiation and a minimum of 50 mJ/cm² (240-280 nm). For a 50

percent transmissive container, the reflector (and lamp) will deliver a minimum of 25 mJ/cm² UV radiation (240-280 nm) to the microorganisms in the container, because 50 % of the energy is absorbed or scattered by the container. Increasing the fluence from the reflectors focused onto the containers, will lower the required energy from the flash lamp, and result in
5 an increase in the lamp life. Further, we determined that the capacitors will need to be upgraded to increase the size of the capacitor to between from 80 and 160 microFarads to produce more energy to increase the amount of UV radiation delivered to the container. The power supply will generate a potential of 2500-6000 volts. The energy will be discharged from the radiation source within 1 millisecond. The reflectors will have enhanced reflection
10 in the ultraviolet. In a preferred embodiment, the reflectors will minimize the non-ultraviolet radiation reaching the medical device. The containers will be located in the focal plane of the reflectors where there is maximum flux.

The criticality that at least a portion of the radiation delivered to the medical device be UV radiation was proven by exposing microorganisms to pulses of a broad spectrum of light
15 (200-1100 nm), and as well as selected portions of the spectrum. Filters were utilized to separate the light spectrum (e.g. ultraviolet: 200 - 400 nm, visible: 400 – 700 nm, infrared:
700 – 1100 nm and full spectrum: 200 – 1100 nm). In this experiment 10⁶ cfu/container of
Aspergillus niger (ATCC 16404) (Figure 3), Bacillus stearothermophilus (ATCC 7953)
20 (Figure 4), and Pseudomonas aeruginosa (ATCC 9027) (Figure 5) were evaluated in a container having a 50% UV transmissive lidstock (240-280 nm) and bowl, and containing non-preserved solution. After inoculation, sample sets of 12 containers of each
25 microorganism were flashed with two 3.5 J/cm² broad spectrum lamp systems with increasing amounts of energy and with a range of filters, and no filter. After the samples were flashed, the samples were diluted and inoculated into an agar medium in which to count the colony forming units. The results are depicted in Figures 3-5. Figure 3 shows the sensitivity of
Aspergillus niger (ATCC 16404) to the different regions of the light spectrum. Figure 4 shows the sensitivity of Bacillus stearothermophilus (ATCC 7953) to the different regions of the light spectrum. In Figure 4, the lines for the IR and VIS overlap. Figure 5 shows the sensitivity of Pseudomonas aeruginosa (ATCC 9027) to the different regions of the light
30 spectrum. In Figure 5, the lines for the UV and Full overlap. The results show that the predominate spectral region for inactivation is the ultraviolet radiation (200-400 nm). Pseudomonas aeruginosa (ATCC 9027) was the microorganism most susceptible in all spectral regions.

An experiment was conducted to determine the sterilization efficacy as a function of radiation intensity, in which fractional positives were obtained at lower energy levels. In separate studies 10^4 cfu/pkg of Bacillus subtilis (ATCC 9372), Bacillus stearothermophilus (ATCC 7953), and Aspergillus niger (ATCC 16404) were added to a container having a 50 %

5 UV-transmissive lidstock and bowl (240-280 nm) with UV-blocking contact lenses in non-preserved solutions. The inoculated containers were exposed to ultraviolet radiation (240 – 280 nm) from 10.2 to 48.6 mJ/cm²/pulse per lamp for 1, 2 and 3 pulses. The system comprised two lamps flashing simultaneously. After exposure the entire contents of the container (contact lens and non-preserved solution) were added to growth medium and

10 incubated at optimal growth temperatures for the test microorganisms. After 14 days incubation, the number of tubes for each set of twelve tubes having viable test microorganism was determined, and recorded in Table 3. Table 3 shows that the threshold of complete cell inactivation for Bacillus subtilis (ATCC 9372) is greater than 18.1 mJ/cm², for Bacillus stearothermophilus (ATCC 7953), it is greater than 30.7 mJ/cm², and for Aspergillus niger

15 (ATCC 16404), it is greater than 18.1 mJ/cm².

TABLE 3

Test Micro-Organism	No. of Flashes	UV radiation (240-280 nm) in mJ/cm ² from each lamp									
		10.2	12.5	15.1	18.1	21.8	26.1	30.7	36.0	41.1	48.6
B. sub.	1	11	7	4	4	0	0	0	0	1	0
	2	0	0	0	0	0	0	0	0	0	0
	3	0	0	0	0	0	0	0	0	0	0
B. stear.	1	12	12	12	12	6	9	4	0	0	0
	2	12	12	12	10	0	0	0	0	0	0
	3	12	12	2	0	0	0	0	0	0	0
<u>Aspergillus niger</u> (ATCC 16404)	1	12	12	8	2	0	0	0	0	0	0
	2	8	2	1	0	0	0	0	0	0	0
	3	1	0	0	1	0	0	0	0	0	0

Figure 6 shows the graph of the average number of containers with Bacillus stearothermophilus (ATCC 7953) survivors in three sets of three containers tested at various UV radiation (240-280 nm) energy levels in mJ/cm² from 12 to 40 mJ/cm² for the fractional data. For the final exposure at 50 mJ/cm² (240-280 nm) three sets of one hundred containers were used, and the average number of positives is reported in the table. Due to the linearity of the data, the D_{value} can be determined from the slope of the graph and by extrapolation a calculation can be made to determine a final dose which will render the package sterile at the desired sterility assurance level. As indicated on the graph the D_{value} of Bacillus

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stearothermophilus (ATCC 7953) is 7.8 mJ/cm² to the container. The container had 50% transmissivity to UV radiation (240-280 nm); therefore, the D_{value} is 3.9 mJ/cm² UV radiation (240-280nm) to the spores for an initial inoculum of 10⁴ spores. For an initial inoculum of 10⁶ spores, the D_{value} of Bacillus stearothermophilus (ATCC 7953) is 7.4 mJ/cm² UV radiation (240-280 nm) to the container, or 3.7 mJ/cm² UV radiation (240-280nm) to the spores. This data was generated using the pair of 3.5 J/cm² broad spectrum lamps, pulsing simultaneously. The additional vertical lines on Figure 6 indicate the amount of UV radiation required to provide a sterility assurance level of 10⁻⁶ for an initial inoculum of 10⁶ and 10⁴ spores.

Although not preferred modes of this invention, it has been determined that this method of sterilization can be enhanced by adding chemicals, bactericides, surfactants, preservatives or heat to the contents of the container. Also the method of sterilization can be enhanced by shaking the container, or exposing the container to sonic vibration energy as it is exposed or between exposures to the radiation source or by heating the container.

This invention has been described with reference to particular embodiments; however, variations within the scope of the following claims are apparent to those of ordinary skill in the art.